

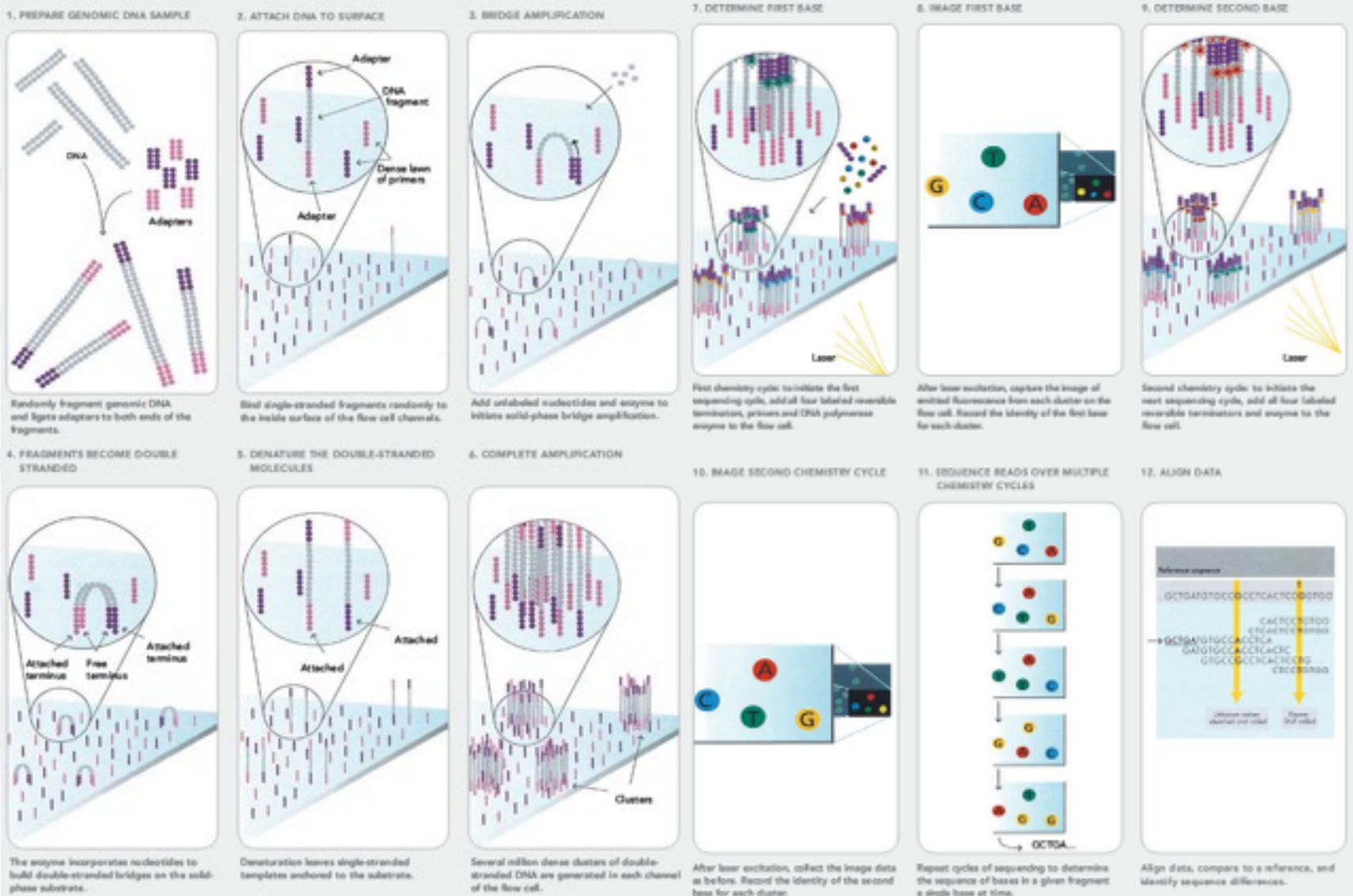


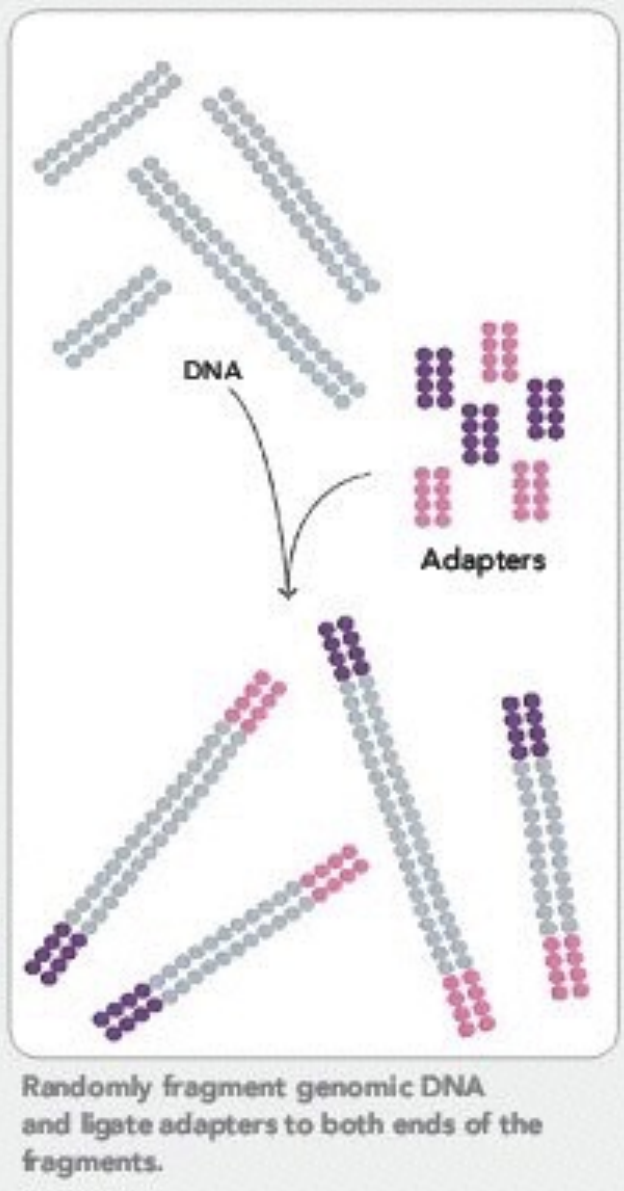
Next Generation Sequencing

Illumina Sequencing

NOTE: These slides are taken from
<http://www.slideshare.net/USDBioinformatics/illumina-sequencing>

Illumina Diagram



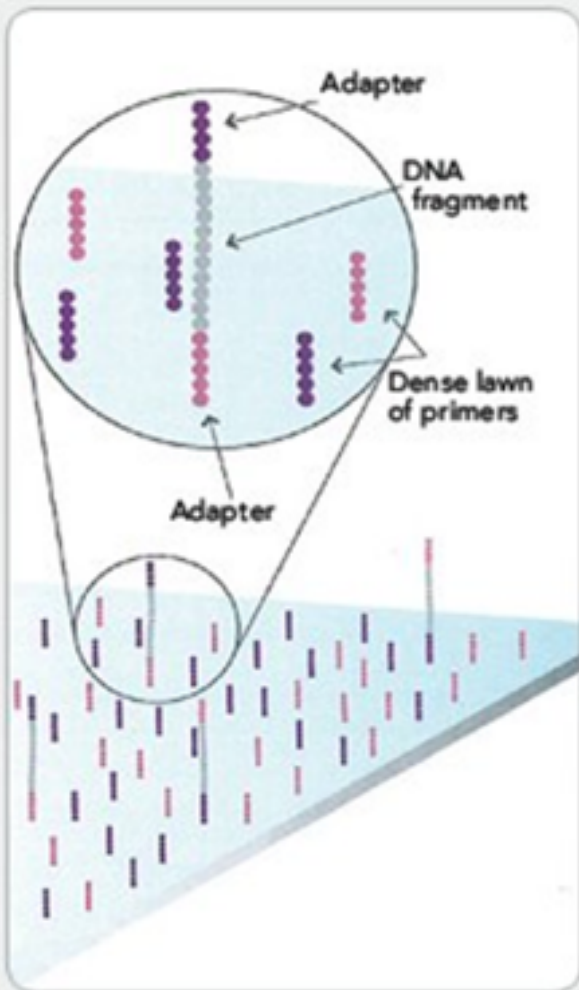


Prepare Genomic DNA Sample



- Fragment DNA of interest into smaller strands that are able to be sequenced
 - Sonication
 - Nebulization
 - Enzyme digestion
- Ligate Adapters
- Denature dsDNA into ssDNA by heating to 95° C

2. ATTACH DNA TO SURFACE



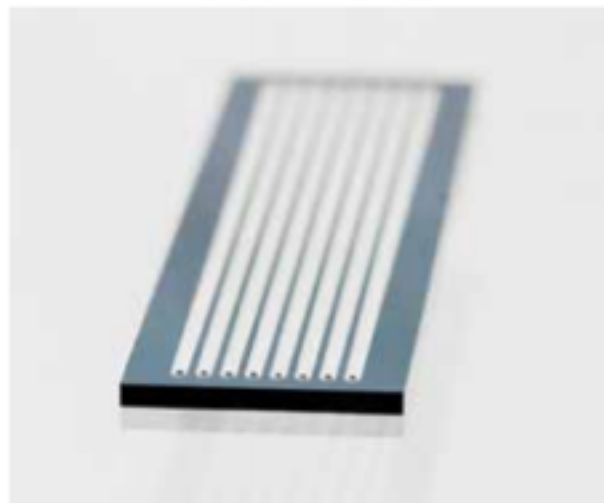
Bind single-stranded fragments randomly to the inside surface of the flow cell channels.

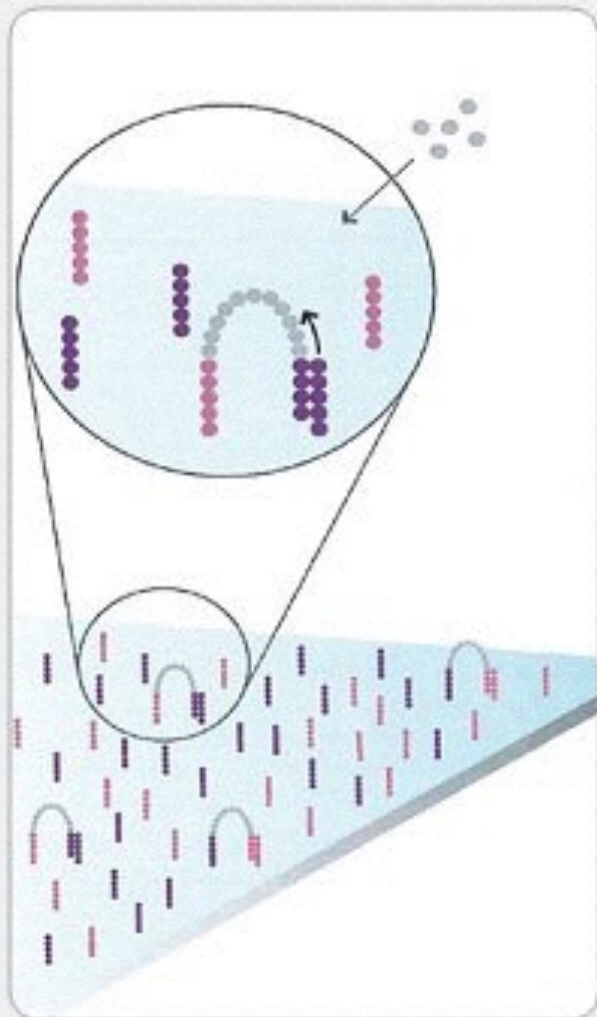
Attach DNA to Surface



- ssDNA is then bound to inside surface of flow cell channels
- Dense lawn of primer on the surface of the flow cell

Flow Cell





Add unlabeled nucleotides and enzyme to initiate solid-phase bridge amplification.

Bridge Amplification



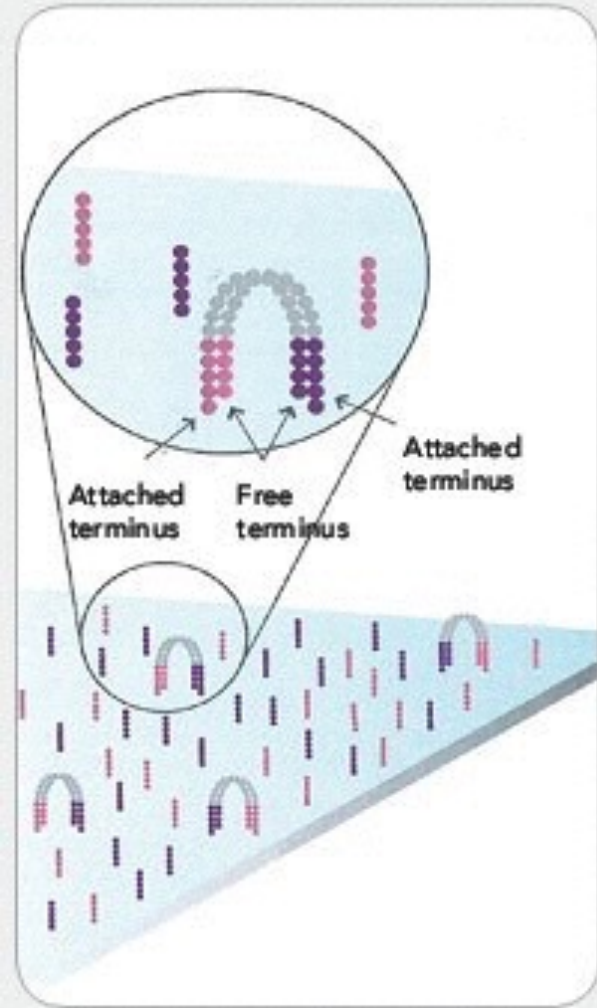
- Unlabeled nucleotides and polymerase enzyme are added to initiate the solid phase bridge amplification

Fragments Become Double Stranded



- In this step it demonstrates the work done by the sequencing reagents
 - Primers
 - Nucleotides
 - Polymerase enzymes
 - Buffer

4. FRAGMENTS BECOME DOUBLE STRANDED

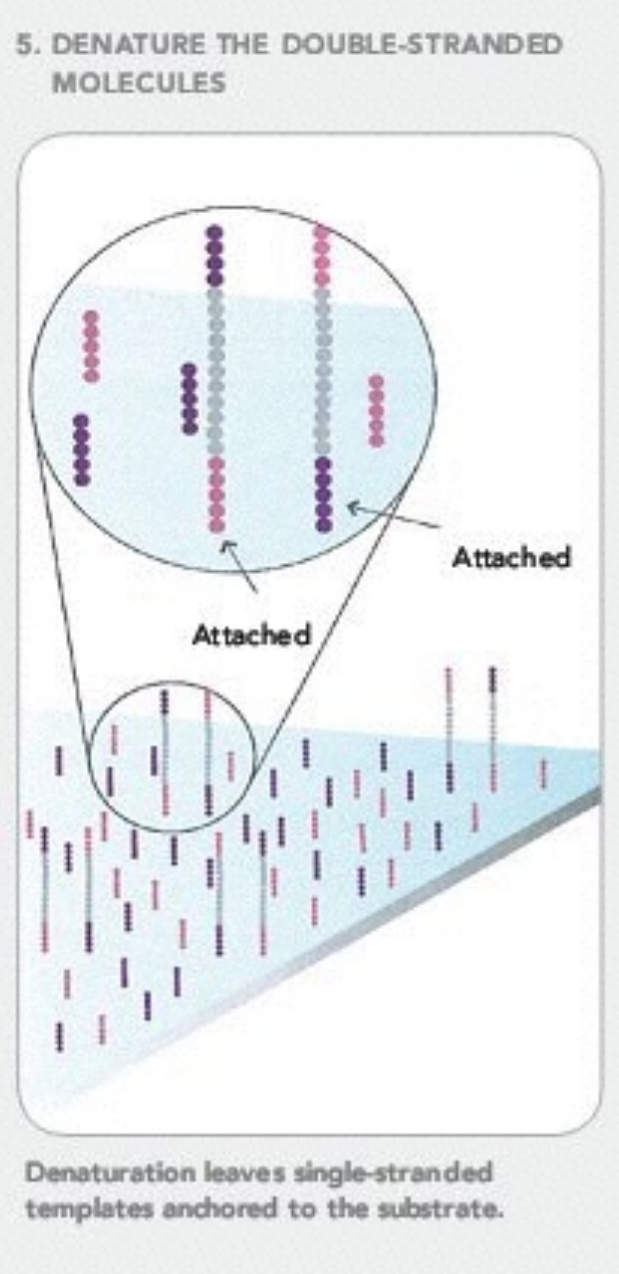


The enzyme incorporates nucleotides to build double-stranded bridges on the solid-phase substrate.

Denature the Double Stranded Molecules



- The original strand is then washed away, leaving only the strands that had been synthesized to the oligos attached to the flow cell



Steps 5-7 Repeats



- Cycle of new strand synthesis and Denaturation to make multiple copies of the same sequence (amplification)
 - Fragments Become Double Stranded
 - Denature the Double Strand Molecules

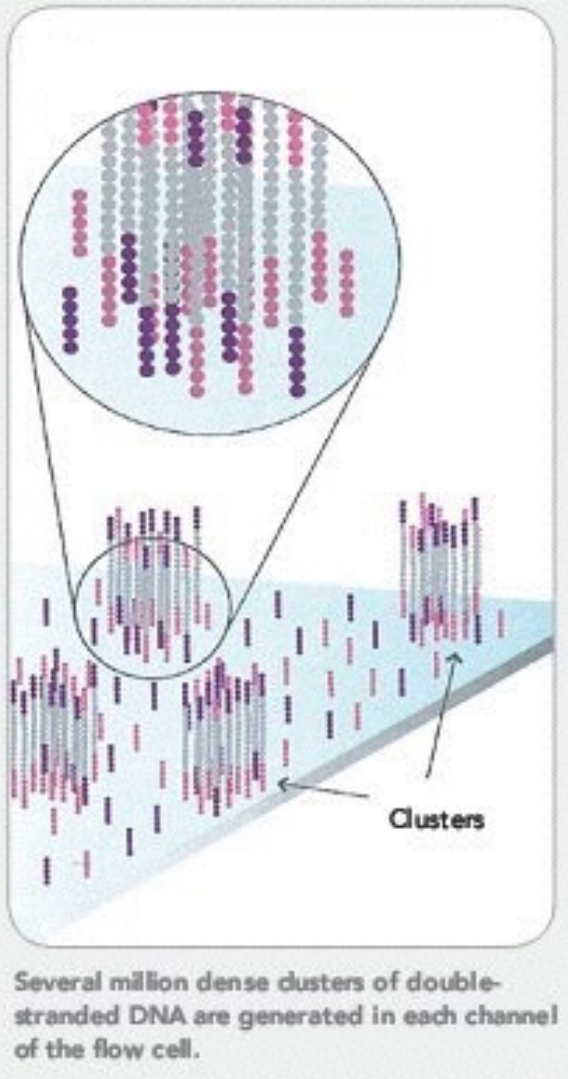
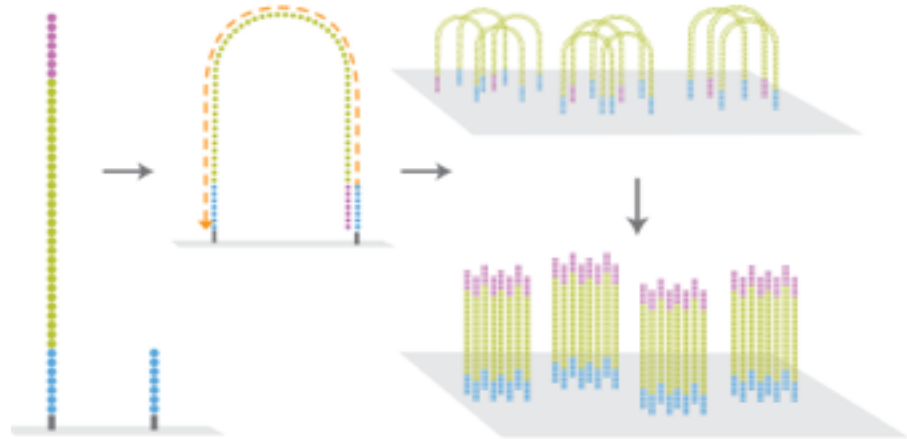
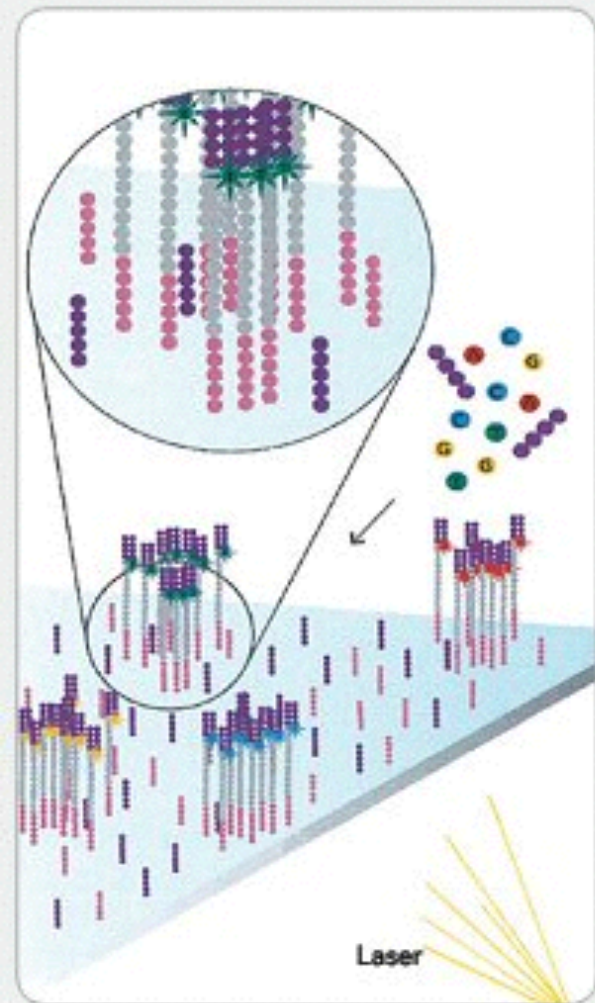


Image retrieved from http://res.illumina.com/documents/products/techspotlights/techspotlight_sequencing.pdf



Determine First Base



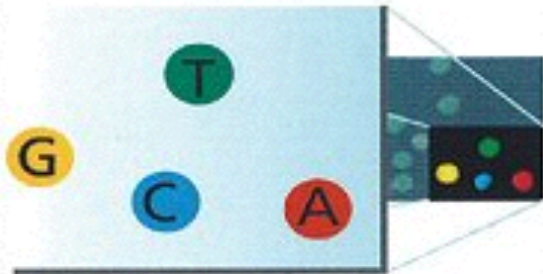
First chemistry cycle: to initiate the first sequencing cycle, add all four labeled reversible terminators, primers and DNA polymerase enzyme to the flow cell.

- The P5 region is cleaved
- Add sequencing reagents
 - Primers
 - Polymerase
 - Fluorescently labeled nucleotides
 - Buffer
- First base incorporated



Image First Base

- Remove unincorporated bases
- Detect Signal
- Deblock and remove the fluorescent signal → new cycle



After laser excitation, capture the image of emitted fluorescence from each cluster on the flow cell. Record the identity of the first base for each cluster.

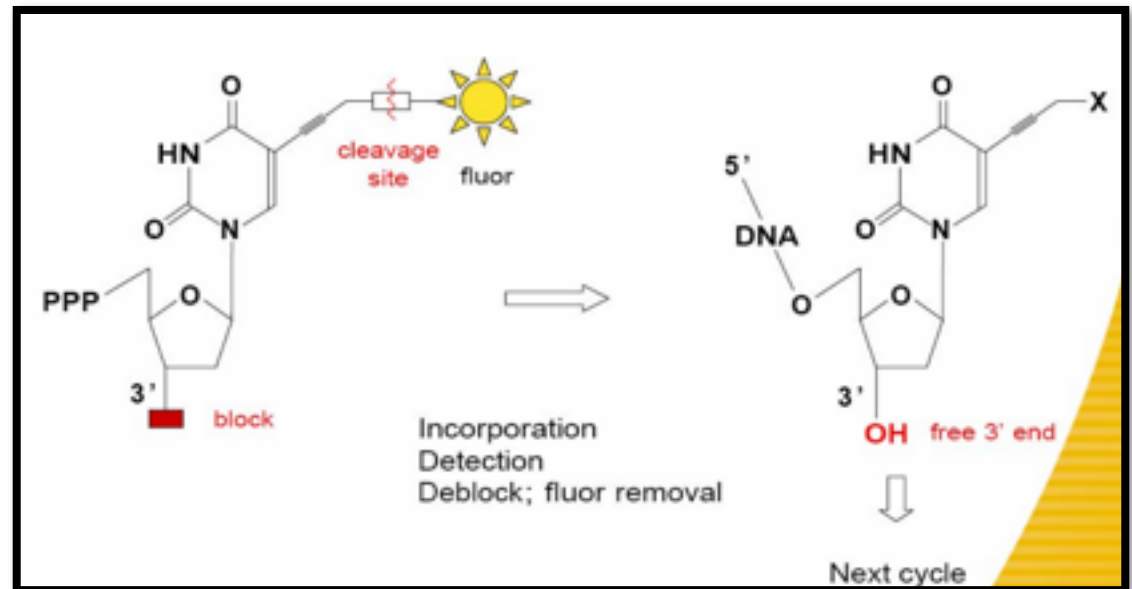
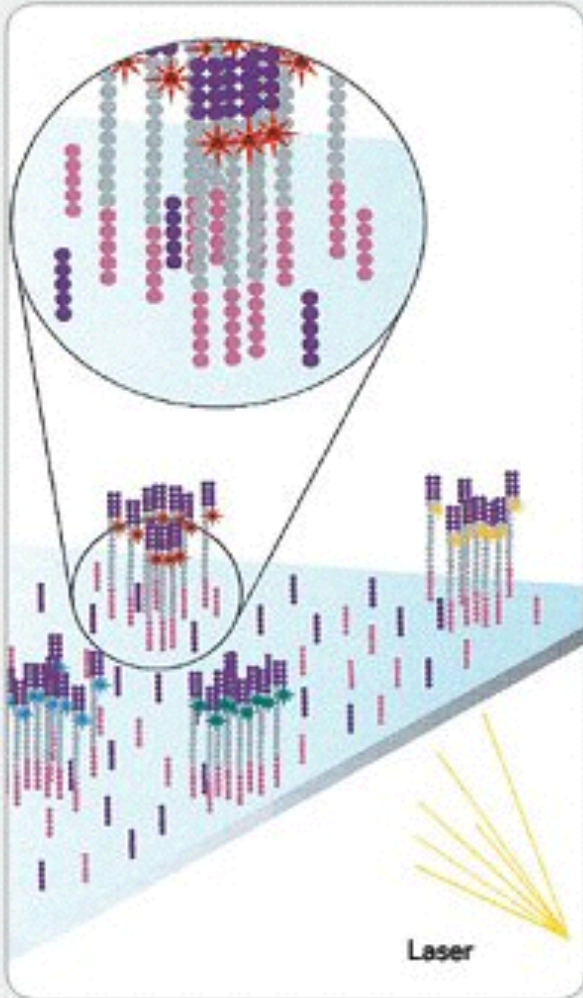


Image retrieved from http://res.illumina.com/documents/products/techspotlights/techspotlight_sequencing.pdf

Image retrieved from http://research.stowers-institute.org/microscopy/external/PowerpointPresentations/ppt/Methods_Technology/KSH_Tech&Methods_012808Final.pdf

Determine Second Base



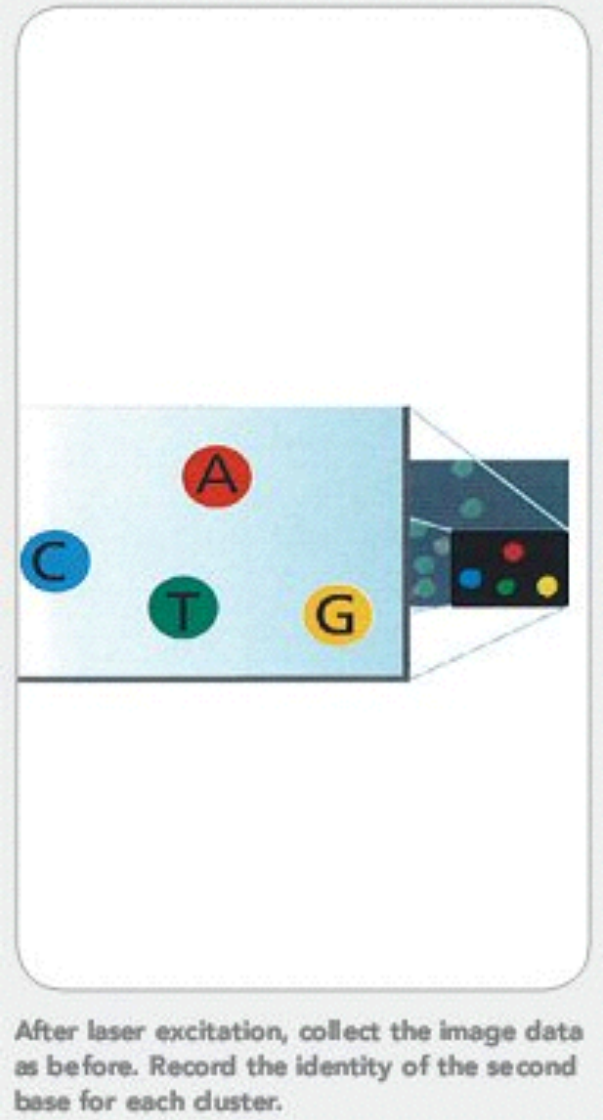
Second chemistry cycle: to initiate the next sequencing cycle, add all four labeled reversible terminators and enzyme to the flow cell.

- Add sequencing reagents
 - Primers
 - Polymerase
 - Fluorescently labeled nucleotides
 - Buffer
- Second base incorporated

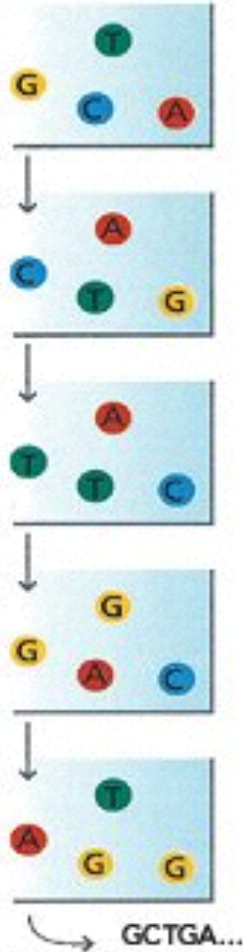
Image Second Chemistry Cycle



- Remove unincorporated bases
- Detect Signal
- Deblock and remove the fluorescent signal → new cycle



11. SEQUENCE READS OVER MULTIPLE CHEMISTRY CYCLES

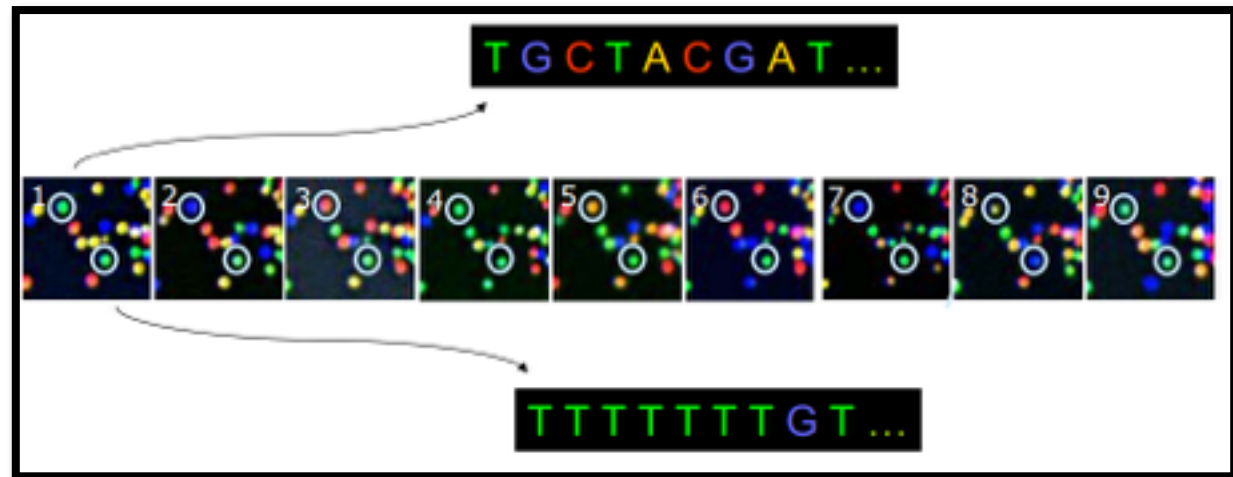


Repeat cycles of sequencing to determine the sequence of bases in a given fragment a single base at time.

Sequence Reads Over Multiple Chemistry Cycles



- The identity of each base of a cluster is read off from sequential images



Work Cited



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